

## Pharmacokinetic variability of in vivo generated <sup>213</sup>Bi and vector labeled <sup>225</sup>Ac in murine cancer models

Since the approval of Xofigo® against metastatic castrated resistant prostate cancer there has been a growing interest in  $\alpha$ -particle emitting radiopharmaceutical therapy. Actinium-225 ( $T_{1/2}=10$ days) is one  $\alpha$ -particle emitter of interest with a total emission of four  $\alpha$ -particles in its decay chain (<sup>221</sup>Fr:  $T_{1/2}=4.9$ min; <sup>217</sup>At:  $T_{1/2}=32.3$ ms; <sup>213</sup>Bi:  $T_{1/2}=45.6$ min; <sup>213</sup>Po:  $T_{1/2}=4.2\mu$ s). Using a chelator makes it possible to label <sup>225</sup>Ac with different vectors, which can be used against different cancers. The bond between the chelator and the <sup>225</sup>Ac atom is relatively weak ( $\sim 10$ eV) and is broken when the  $\alpha$ -particle is emitted from the <sup>225</sup>Ac atom due to the high recoil energy ( $\sim 100$ keV). Consequently the <sup>225</sup>Ac decay daughters are not bound to the vector and are free to relocate. Depending on the vector's pharmacokinetics there will be different normal tissue uptake of vector labeled <sup>225</sup>Ac. This suggests that depending on vector type the unbound decay daughters are present in different tissues at different concentrations, able to redistribute within the *in vivo* system.

In this study we investigated vector labeled <sup>225</sup>Ac and unbound <sup>213</sup>Bi, which has the longest half-life of the <sup>225</sup>Ac decay daughters and is known to relocate and accumulate in the kidneys. We compared the pharmacokinetics of *in vivo* generated <sup>213</sup>Bi and vector labeled <sup>225</sup>Ac for three different vectors in murine cancer models (7.16.4 antibodies against breast cancer; anti-VLA-4 antibodies against melanoma; PSMA-targeted small molecule against castrate resistant prostate cancer). The mice were injected i.v. by the tail vein with the <sup>225</sup>Ac labeled vectors and sacrificed at specific times after administration. The organs and tissues of interest were blood, liver, kidneys and spleen, which were harvested and directly measured in a gamma well counter in 1 minute intervals for up to 5 hours. The measured data was fitted with a bi-exponential function to determine the amount of unbound <sup>213</sup>Bi and vector labeled <sup>225</sup>Ac present in the different tissues.

All murine models showed accumulation of unbound <sup>213</sup>Bi in the kidneys, accounting for  $\sim 60\%$  of the mean absorbed dose to the kidneys. The main supplier of the unbound <sup>213</sup>Bi to the kidneys for the small molecule and the anti-VLA-4 was the liver, and for the 7.16.4 the blood. In addition, vector labeled <sup>225</sup>Ac has shown to be uniformly distributed for 7.16.4, and non-uniform for anti-VLA-4, PSMA-targeting small molecule and unbound <sup>213</sup>Bi within the kidneys, suggesting the need for small scale dosimetry for an accurate absorbed dose calculation.

This study shows that the delivery of <sup>225</sup>Ac using different vectors changes the pharmacokinetics and supply of unbound <sup>213</sup>Bi to the kidneys. This is important for translation to clinical studies as *in vivo* imaging of <sup>225</sup>Ac does not distinguish between the vector labeled <sup>225</sup>Ac and unbound <sup>213</sup>Bi, resulting in an possible overestimation of the absorbed dose to the kidneys and underestimation of the supplying tissues.

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